

OVERVIEW OF SULFUR IN PLANT NUTRITION

Sulfur (S) is an essential nutrient for plant growth. It is considered a macronutrient and must be available in relatively large amounts for good crop growth—much like nitrogen (N), phosphorus (P), potassium (K), calcium (Ca), and magnesium (Mg). Sulfur is present in plants as part of the amino acids cysteine, cystine, and methionine, which make proteins. These amino acids account for about 90% of the S in plants. Thus, protein synthesis and photosynthetic rates are decreased when S is deficient. Cysteine and methionine are also precursors of other S-containing compounds, such as coenzymes and secondary plant products. Sulfur is part of the structure of these compounds or acts as a functional group directly involved in metabolic reactions. Plants mainly absorb S in the form of sulfate ions (SO_4^{-2}) (Mengel and Kirkby, 1982).

Nationwide, deficiencies of S in crops are thought to be increasing (McGrath and Zhao, 1995). Sulfur deficiencies are attributed to improved fertilizers that contain little to no S impurities, intensive cropping systems that leave behind little organic matter, increased yield that results in more S removal, less deposition of S from the atmosphere, and less use of S-containing pesticides.

Sulfur-deficient soils are often low in organic matter, coarse textured, well drained, and subject to leaching because sulfate is mobile in the soil. In semi-arid regions, SO_4^{-2} can accumulate in the lower soil profile as soluble gypsum. The S status of New Mexico's soils is not well defined, and S effects on the growth of New Mexico crops have not been extensively researched.

PLANT SULFUR NEEDS

Sulfur deficiency symptoms are sometimes difficult to distinguish from N deficiency. However, in contrast to N deficiency, S deficiency symptoms first occur in the younger, most recently developed leaves. Plants

that are S-deficient have a reduced rate of growth and can be rigid and brittle, and the stems remain relatively thin. These plants are also uniformly chlorotic (yellowish color or lacking chlorophyll).

Diagnosing S deficiency requires proper plant tissue analyses. Gavlak et al. (1994) recommend a 2% (v/v) acetic acid extraction for New Mexico and other western states. Plant leaves can be tested for organic N and organic S content. A high ratio of organic N to organic S (70:1–80:1), as compared to healthy tissue (<50:1), is considered a diagnostic test since the S-containing amino acids are not formed in S-deficient plants. Another diagnostic tool for confirming S-deficient plants is an accumulation of nitrate-N. Plants with lower total protein and sugars in the leaf blades and stems can have a surplus of nitrate-N in the tissue (Ergle and Eaton, 1951).

Plant tissue levels for total S have different minimum levels for optimum production. Wheat, sunflower, and field beans have some of the highest requirements, whereas corn and soybeans have some of the lowest critical levels (Hitsuda et al., 2005) (Table 1). Plant tissue analysis interpretations should reflect these differences among crops. Mathot et al. (2009) have also recently proposed three diagnostic zones for total S and total N content for determining if grasses are S-deficient. Their guidelines include the effects of N nutrition and the dilution of the grass S content during growth. Grasses are S-deficient, in their proposed diagnostic tool, when the total S content (mg/g) falls on or below a line described by the equation $0.0665 \times \text{N} - 0.2805$ (total N is also presented in mg/g) (Figure 1). Validation of this tool is needed for New Mexico. Please share your results with your local county Extension agriculture agent.

The S content of plants is approximately the same as the P content (Mengel and Kirkby, 1982). However, S fertilization is not as critical as P since SO_4^{-2} is much more mobile and not as strongly fixed to soil particles

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Table 1. Critical Plant S Concentration for S Deficiency with 75% Relative Shoot Dry Weight Compared to Optimum Plant Growth (adapted from Hitsuda et al., 2005, and as referenced by Dick et al., 2008)

Crop	Plant part and growth stage	Shoot S† concentration	
		Critical growth mg g ⁻¹	Optimum growth mg g ⁻¹
Alfalfa (<i>Medicago sativa</i> L.)	Top 6 in. at early bud	2.0	2.6–5.0
Corn (<i>Zea mays</i> L.)	Ear leaf at silking	1.0	2.1–5.0
Corn	Shoot 30 days after emergence	0.76	2.06
Cotton (<i>Gossypium hirsutum</i> L.)	Youngest mature leaf blade at early flower	<2.0	2.0–2.5
Cotton	Shoot 30 days after emergence	1.07	4.1
Field bean (<i>Phaseolus vulgaris</i> L.)	Shoot 30 days after emergence	1.56	2.56
Oat (<i>Avena sativa</i> L.)	Top leaves at boot stage	<1.5	2.1–4.0
Sorghum (<i>Sorghum bicolor</i> (L.) Moench)	Shoot 30 days after emergence	1.13	1.99
Peanut (<i>Arachis hypogaea</i> L.)	Youngest mature leaf at preflower	<2.0	2.0–3.5
Sunflower (<i>Helianthus annuus</i> L.)	Shoot 30 days after emergence	1.56	3.12
Wheat (<i>Triticum aestivum</i> L.)	Shoot 30 days after emergence	1.43	2.33

†Determined by spectrometer after wet washing dried tissue with nitric and perchloric acid.

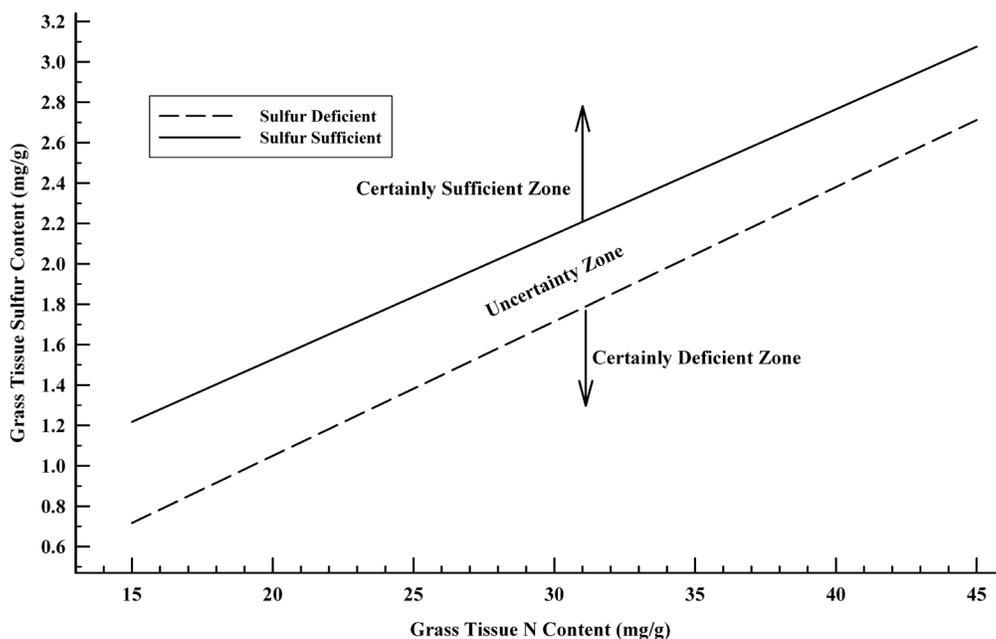


Figure 1. Boundary lines for identifying S sufficiency and S deficiency in grasses when given grass tissue total N and total S content in mg/g (adapted from Mathot et al., 2009).

Table 2. Sulfate Concentration in Selected Waters Used for Irrigation

Location	mg/L SO ₄ ⁻²	lb/acre inch
Otero County	1,921	427
San Miguel County	5	1.1
Lea County	767	170
Doña Ana County	1,440	320
Eddy County	1,100	244

as P. Alfalfa, a major crop in New Mexico, has a relatively high S requirement. Approximately 5.3 lb of S are removed for every ton of harvested alfalfa (Chen et al., 2005). A ten ton yield goal for the Doña Ana County region of New Mexico would be expected to remove 50 to 55 lb/ac of S. Westermann (1975) reported that the critical biomass value for alfalfa is 0.15 to 0.20% S. Cotton lint can remove nearly 53 lb S/ac with a 3.2 bale per acre yield (Dick et al., 2008). The youngest, most fully developed leaf of cotton at early flowering should have between 0.20 and 0.25% S to be considered sufficient.

SULFATE SOURCES IN NEW MEXICO

Soil

Plants derive sulfate from soil solution SO₄⁻² and adsorbed inorganic SO₄⁻² (Kowalenko and Grimmett, 2008). Inorganic sulfate may be present in soil water, bound or adsorbed on soil surfaces, bound to gypsum (Nelson, 1982), or associated with calcium carbonate (Roberts and Bettany, 1985). Determining plant-available SO₄⁻², along with the plant-available N, in the soil can give the grower a “snapshot” of whether or not additional SO₄⁻² is needed for plant growth. Soils are generally considered sufficient in SO₄⁻² if the ratio of plant-available SO₄⁻² to plant-available N is less than 15:1.

There are a host of soil extracts that can be used to help determine plant-available SO₄⁻², including acetates, carbonates, chlorides, citrates, and oxalates. The preferred extraction method uses calcium phosphate (Beaton et al., 1968), which is used by the NMSU Soil Water Agricultural Testing laboratory. The phosphate ion is used to displace sulfate on soil adsorption sites. Many New Mexico soils, however, fix considerable amounts of phosphate, and labs must compensate by increasing the phosphate concentration in the extract. This “snapshot” does not account for sulfate that may become available from organic matter or other soil inputs or sources. It also does not account for what can happen to sulfate during the growing season as it cycles through the soil, plant, and air systems.

Sulfur cycling (Figure 2) in soils is a complex and dynamic process as outlined by Eriksen (2008). Inputs

to soil solution SO₄⁻² include atmospheric S, animal manures, fertilizers, and decomposition of organic sources. Sulfate-S from the soil solution can be absorbed by the plant, adsorbed to soil particles, converted into other stable inorganic compounds, or leached beyond the plant root zone. Attention to irrigation water management, however, is important for SO₄⁻² management since SO₄⁻² is easily leached through many New Mexico soils, particularly sandy or coarse-textured soils. It is also recommended that soils be sampled below the plow layer since SO₄⁻² can be stored at lower depths in the soil profile due to leaching.

Irrigation Water

Irrigation water can be a major source of S for New Mexico fields. Table 2 illustrates the SO₄⁻²-S variability from selected locations in New Mexico. An irrigation water sample with 4.5 mg/L SO₄⁻² would add approximately 1 lb SO₄⁻² per acre inch of applied water.

Gypsum

Calcium sulfate (CaSO₄·2H₂O), or gypsum, is slightly soluble in water (~2 g/L = 8.3 lb per 1,000 gal; National Institute of Occupational Safety and Health, 2003) and can therefore provide some sulfate for plants. Many soils in New Mexico are gypsiferous (>2% calcium sulfate, approximately 76,134 lb gypsum per acre) and are usually not prone to S deficiency. A soil with at least 2% gypsum should have at least 107 lb of SO₄⁻² available for uptake under adequately watered conditions. Gypsum is used primarily to reclaim sodium (Na)-affected soils to provide the Ca needed to displace Na. A side benefit can be the SO₄⁻² for plant uptake. However, SO₄⁻² is a soluble anion that will contribute to soil salinity.

Elemental Sulfur

Oxidation of elemental sulfur (S⁰) to plant-available SO₄⁻²-S is mediated by soil microorganisms, primarily *Thiobacillus* spp. Oxidation of sulfur is influenced by its particle size, as well as the soil's temperature and moisture content. Warm, moist soil conditions are needed for optimal biological oxidation to occur, but oxidation will occur slowly in cool, moist soils. Additionally, the rate of S⁰ oxidation generally increases as the organic matter content increases. Several S⁰ sources have been used in New Mexico in an effort to affect soil pH, micronutrient availability, and SO₄⁻²-S content. Lindemann et al. (1991) evaluated several S sources, including reagent-grade S⁰, wettable S (90% S⁰), Disper-Sul S (90% S⁰), and flowable S (52% S⁰), on calcareous soils. The small particle size (<425 μm) of the flowable S was more effective than the other sources of S⁰ when tested in the field, but only decreased pH by 0.1 to 0.2 units, which probably had no biological significance. However, the flowable form kept the pH at a lower level for more

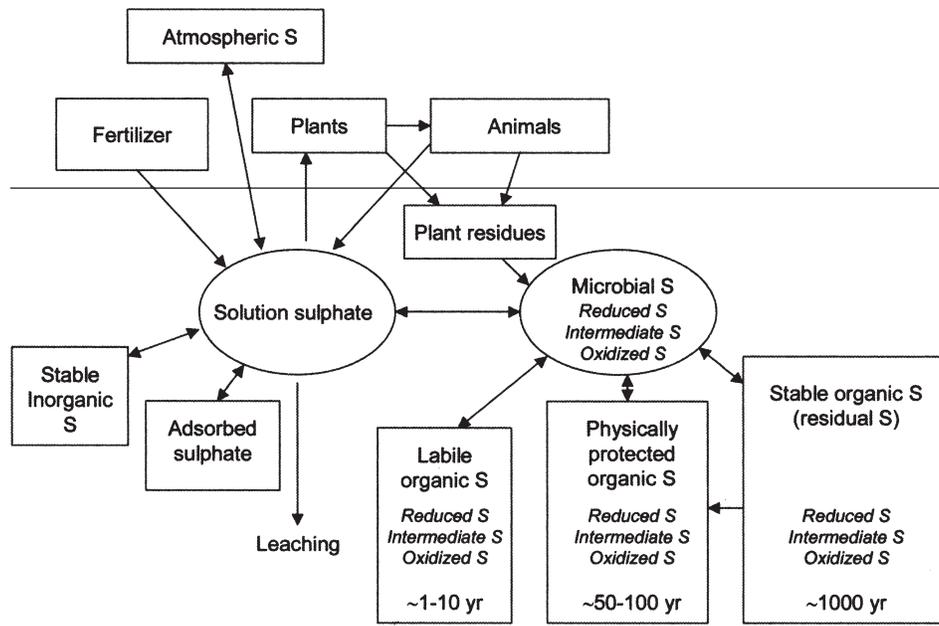


Figure 2. Conceptual model that demonstrates the soil sulfur cycle (from Eriksen, 2008; used with permission).

than 12 weeks after application, which was significantly longer than all other S sources. Wetttable S increased soil salinity by 2 mmhos/cm, which could be harmful to salt-sensitive plants. The SO_4^{-2} content increased by 1,000 ppm (mg/kg) as a result of using any of the tested S sources. This level of SO_4^{-2} gradually decreased as the soil was irrigated and the SO_4^{-2} leached from the root zone.

Organic Matter

The addition of organic matter can improve soil SO_4^{-2} -S content, depending on the composition and source of the organic matter (Cifuentes and Lindemann, 1993). When added to a clay soil, fresh cow manure (0.30% S) or bermudagrass clippings (0.22% S) resulted in 63.5% more soil SO_4^{-2} than composted horse manure (0.10% S) during the first 8 weeks after application. The difference gradually decreased to just 9.4%, probably reflecting a loss of SO_4^{-2} by leaching. Greater S^0 oxidation in the presence of fresh cow manure or bermudagrass clippings was attributed to their higher N content and their ease of decomposition, which allowed for more decomposition of the S-containing carbon compounds. The higher S content and faster decomposition probably promoted a larger number of chemoheterotrophic S microbes than the more resistant composted horse manure (Cifuentes and Lindemann, 1993). Generally speaking, manures and plant material should have a relatively high N and S content to promote more plant-available SO_4^{-2} . It is difficult, however, to predict how much plant-available SO_4^{-2} can come from added organic sources of S because of

complicated dynamics in the soil environment (Eriksen, 2008; Scherer, 2001).

Other Sulfate Amendments

Many methods are available to increase soil organic matter, including green manures (turning cover crops into the soil), reduced tillage or no-tillage, and adding organic matter from farmyard manures.

Sulfate-containing fertilizers include, but are not limited to,

- ammonium sulfate (24% S),
- ammonium thiosulfate (26% S),
- ordinary superphosphate (11–12% S),
- magnesium sulfate (14% S),
- sulfate of potash magnesia (22% S), and
- potassium sulfate (18% S).

SULFATES AND GROUNDWATER QUALITY

Ground and surface water quality are regulated under the authority of chapter 6, part 2 of New Mexico's Title 20 environmental protection regulations (New Mexico Administrative Code, 2001). The standards for groundwater with less than 10,000 mg/L of total dissolved solids (TDS) limit the SO_4^{-2} concentration for domestic water supplies to less than 600 mg/L unless the existing condition (natural state) exceeds that standard. Additionally, domestic water supplies must not exceed 1,000 mg TDS/L.

Table 3. Sulfate Levels in Drinking Water for Cattle

Sulfate level (mg/L)	Interpretation
<500	Safe for drinking
500–1,500	Generally safe; trace mineral availability may be reduced; may decrease performance in confined cattle
1,500–3,000	Marginal; may be unsuitable for confined cattle during hot weather; performance may be reduced; sporadic cases of polioencephalomalacia (PEM) may occur
3,000–4,000	Unsuitable; decreased performance of grazing cattle may occur, and risk for PEM in confined cattle is increased
>4,000	Dangerous; health problems expected and substantial reductions in cattle performance; secondary copper deficiency likely

SULFATE AND CATTLE

The National Research Council (1996) gives the daily S requirement for cattle as 0.15% of diet dry matter, and the maximum tolerable concentrations of dietary S are estimated to be 0.40% of dry matter intake. Runyan et al. (2009) and Ellis (2008) offer guidelines for sulfate in drinking water for cattle (Table 3).

BEST MANAGEMENT PRACTICES

Sulfate plays a critical role in plant and animal health in New Mexico. Long-term success in growing plants includes knowing whether or not SO_4^{-2} is needed. A sustainable approach to managing sulfate nutrition includes the following actions.

- Test the soil for plant-available SO_4^{-2} .
- Test the irrigation water as a viable source of SO_4^{-2} for plants.
- Evaluate the plant tissue S content for critical levels at specific growth stages.
- Evaluate crop rotations that conserve or enhance SO_4^{-2} nutrition, such as high residue crops that return SO_4^{-2} sources to the soil or are not heavy SO_4^{-2} users under SO_4^{-2} -limited conditions.
- Evaluate management practices that can build soil organic matter, which would include the responsible use of manures, green manures, and cover crops.
- Manage irrigation water to avoid excess leaching that could reduce soil SO_4^{-2} .
- Consider SO_4^{-2} amendments if all the possible sources and plant requirements do not balance.

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